

Multi-organ-on-chip Device for Modeling Opioid Reinforcement and Withdrawal, and the Negative Affective Component of Pain: a Therapeutic Screening Tool

Wang ZZ¹, Shu Z², Laperle AH³, Ruckodanov D², Kawakita S⁴, Jucaud V⁴, Ashammakhi N², Monbouquette H², Dokmeci MR⁴, Khademhosseini A⁴, Svendsen C³, Seidlits SK¹, Maidment NT²

¹Univeristy of Texas at Austin, Austin; ²University of California Los Angeles; ³Cedars Sinai Los Angeles; ⁴Terasaki Institute for Biomedical Innovation, Los Angeles; of

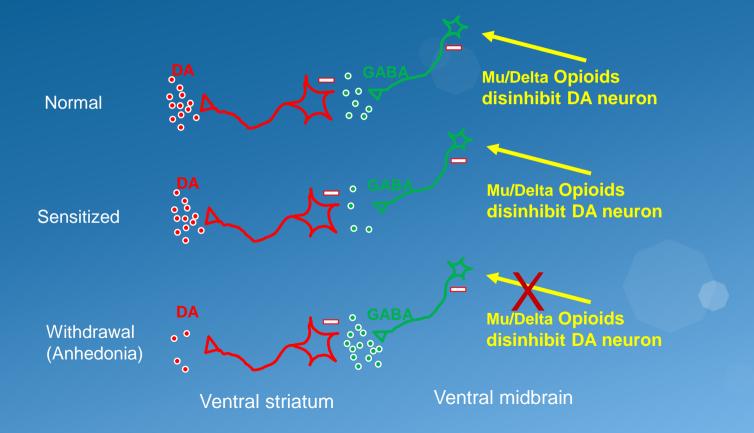


Objective

Design "multi-organ-on-chip" model systems based on human cells and use them to better understand the addictive process at a molecular level and to potentially identify new therapeutics to treat drug addiction and chronic pain.

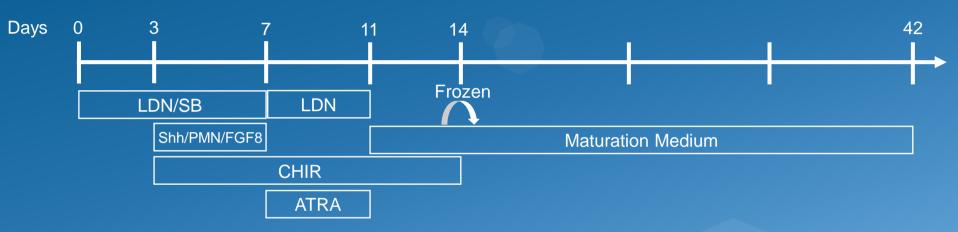


How Opioids Affect Neurons





Midbrain Neuron Differentiation Protocol



iPS Cell Line: Lothian Birth Cohort 1936 iPSC Collection, CS1185iCTR-LBCn2 (EDi044-A)

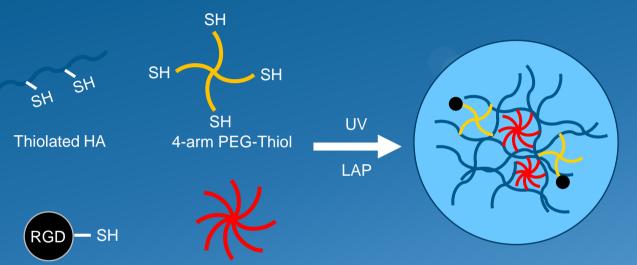
LDN: BMP inhibitor SB: TGF-β inhibitor Shh: Sonic Hedgehog PMN: Purmophamine FGF8: Fibroblast growth factor 8

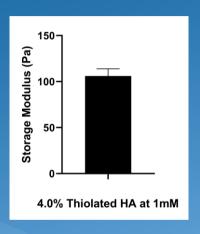
CHIR: GSK3 inhibitor ATRA: All-trans retinoic acid

Maturation Medium: BDNF, GDNF, dbCAMP, Ascorbic acid, DAPT, TGF-β3



HA Hydrogel Fabrication Using Photocrosslinking





Integrin-Binding Peptide 8-arm PEG Norbornene

Storage Modulus = 106.09 ± 19.34 Pa (N = 3, 13 sweeps from each gel)

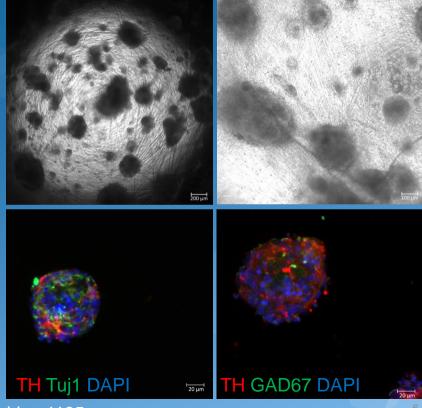
HA: hyaluronic acid, ~700 kDa, 4% thiolation, 0.5wt%

Cysteine-terminated RGD peptides added to ~25% of PEG-thiol arms for cell adhesion.

LAP: type I photoinitiator lithium phenyl(2,4,6-trimethylbenzoyl)phosphinate, 0.025 wt%

UCLA TEXAS STERASAKI Social Singi iPSC-derived Midbrain Populations in HA Hydrogel

- Network formation was observed
- Cryosections of the gel stained positive for midbrain population markers
 - Neuronal marker β-tubulin III (Tuj1)
 - Dopaminergic neuron marker tyrosine hydroxylase (TH)
 - GABAergic neuronal marker glutamate decarboxylase 67 (GAD67)

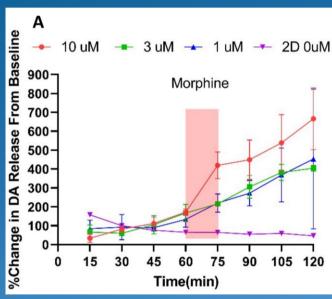


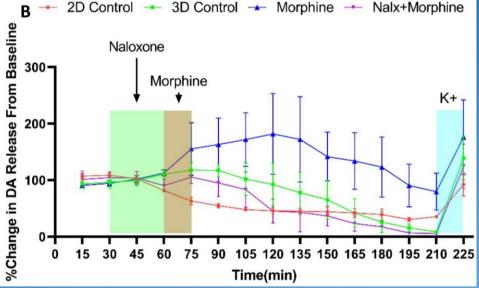
Line 1185

6



Dopamine Release Response to Morphine and Naloxone in 3D Culture

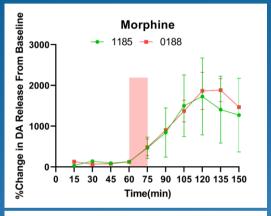


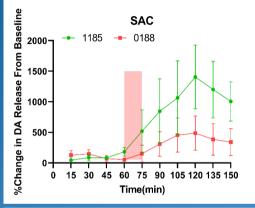


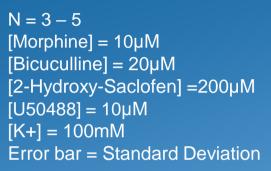
Line 0188 N = 3 Error Bar = Standard Deviation Line 0188
[Morphine] = 10µM; [Naloxone] = 30µM; [K+] = 100 mM
N = 3
Error Bar = Standard Deviation

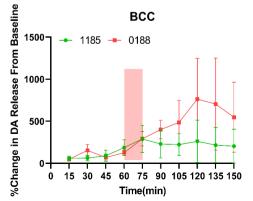


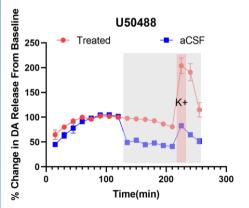
Dopamine release induced by morphine, GABA antagonists, and κopioid agonist





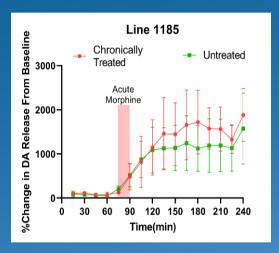




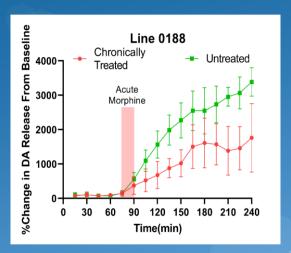




Effect of chronic morphine exposure followed by withdrawal on dopamine release induced by a subsequent morphine challenge



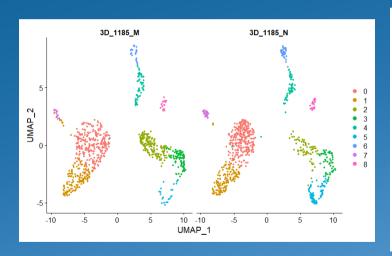
N = 4 - 5[Morphine] = $10\mu M$ Error bar = Standard Deviation

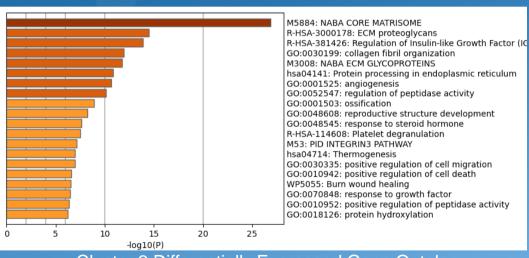


Chronic exposure: 4 h daily, 10 µM morphine, 7 days Withdrawal: additional 7 days before acute morphine exposure



scRNAseq of Chronic Morphine Treatment

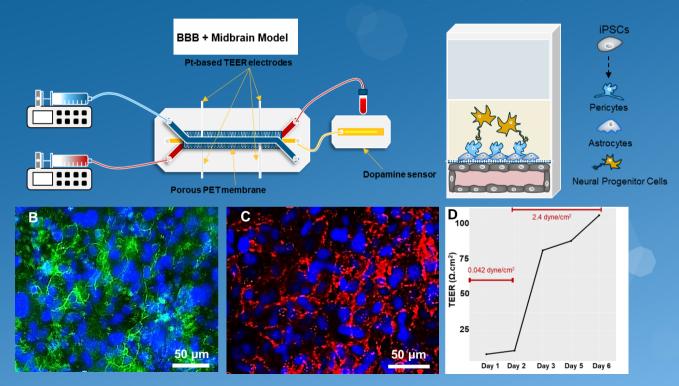




Cluster 2 Differentially Expressed Gene Ontology

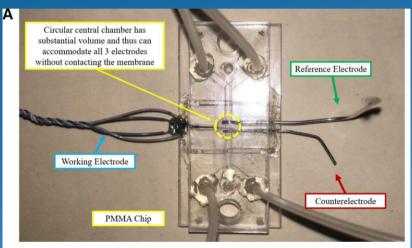


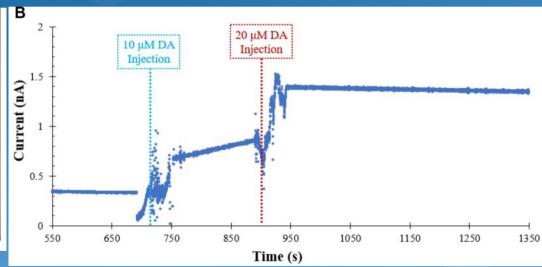
Immortalized brain microvascular endothelial cells (hBMECs) cultured in the BBB chip





Integration of PMMA-based BBB chip with DA microarray sensor







Summary

- Observed reproducible, concentration-dependent, morphine-induced dopamine release from 2D and 3D cultures of two iPSC lines that is blocked by naloxone.
- Exposure to the GABA-B receptor antagonist, 8-OH-saclofen, increases basal efflux of dopamine indicating that dopamine neurons are tonically inhibited by the presence of endogenous GABA, reproducing the in vivo midbrain circuitry.
- Exposure to the kappa opioid receptor agonist, U50488, significantly attenuates basal and K+-induced dopamine efflux.
- Preliminary experiments have provided evidence of plasticity in dopamine release following chronic morphine exposure. The amplitude of the dopamine response to morphine challenge was attenuated in chronically-exposed cultures of line 0188 (male), but not line 1185 (female). Such tolerance in dopamine response potentially models the anhedonic component of withdrawal.
- Observed a substantial effect of shear stress on TEER and achieved values > 100 Ω.cm2.
- Incorporated on-line amperometric dopamine detection with silicon wafer-based electrodes into chip design.

UCLA TEXAS STERASAKI Cocadars Sinai Acknowledgment

Seidlits Lab

Dr. Ze Zhong Wang

Maidment Lab

Dr. Zhan Shu

Svendsen Lab

Dr. Alexander Laperle

Dr. Samuel Sances

Dr. Deepti Lall

Terasaki Institute

Dr. Ali Khademhosseini

Dr. Mehmet Dokmeci

Satoru Kawakita

Monbouquette Lab

Dima Ruckodanov

Dr. Nureddin Ashammakhi

Dr. Cathy Cahill

Dr. Robert Damoiseaux



Technology Center for Genomics & Bioinformatics At UCLA